

*OCA2*481Thr*, a hypofunctional allele in pigmentation, is characteristic of northeastern Asian populations

Isao Yuasa · Kazuo Umetsu · Shinji Harihara · Aya Miyoshi · Naruya Saitou ·
Kyung Sook Park · Bumbein Dashnyam · Feng Jin · Gérard Lucotte ·
Prasanta K. Chattopadhyay · Lotte Henke · Jürgen Henke

Received: 11 April 2007 / Accepted: 24 May 2007 / Published online: 14 June 2007
© The Japan Society of Human Genetics and Springer 2007

Abstract Asians as well as Europeans have light skin, for which no genes to date are known to be responsible. A mutation, Ala481Thr (c.G1559A), in the oculocutaneous albinism type II (*OCA2*) gene has approximately 70% function of the wild type allele in melanogenesis. In this study, the distribution of the mutation was investigated in a total of 2,615 individuals in 20 populations from various areas. *OCA2*481Thr* prevailed almost exclusively in a northeastern part of Asia. The allele frequency was highest in Buryat (0.24) in Mongolia and showed a north–south downward geographical gradient. These findings suggest that *OCA2*481Thr* arose in a region of low ultraviolet radiation and thereafter spread to neighboring populations.

Keywords *OCA2* · Pigmentation · Polymorphism · Population study · Skin color

Introduction

Skin color is a complex genetic trait that has long intrigued biologists. Its variation among indigenous peoples is obvious and correlated with levels of ultraviolet radiation, becoming lighter in more northerly latitudes (Jablonski and Chaplin 2000). Asians as well as Europeans have light skin, for which no genes to date are known to be responsible. Recent interest in signatures of positive selection in candidate pigmentation genes has been raised by a long-range haplotype test and an F_{ST} -based approach (Izagirre et al. 2006; Lao et al. 2007; Myles et al. 2007). Some candidate genes appear to have evolved independently after the divergence of Europeans and East Asians. The *OCA2* gene, showing relatively strong signatures for natural selection, is included among the candidate genes affecting

I. Yuasa (✉)
Division of Legal Medicine, Faculty of Medicine,
Tottori University, Yonago 683-8503, Japan
e-mail: yuasai@grape.med.tottori-u.ac.jp

K. Umetsu
Department of Experimental and Forensic Pathology,
Faculty of Medicine, Yamagata University, Yamagata, Japan

S. Harihara
Department of Biological Sciences,
Graduate School of Science,
University of Tokyo, Tokyo, Japan

A. Miyoshi
Department of Forensic Medicine,
Fukuoka University School of Medicine, Fukuoka, Japan

N. Saitou
Division of Population Genetics,
National Institute of Genetics, Mishima, Japan

K. S. Park
Department of Biology, Sungshin Women's University,
Seoul, South Korea

B. Dashnyam
Institute of Biological Sciences, Mongolian Academy
of Sciences, Ulaan Baator, Mongolia

F. Jin
Institute of Genetics and Developmental Biology,
Chinese Academy of Sciences, Beijing, China

G. Lucotte
Center of Molecular Neurogenetics, Paris, France

P. K. Chattopadhyay
Amity Institute of Forensic Sciences,
Defence Colony, New Delhi, India

L. Henke · J. Henke
Institut für Blutgruppenforschung, Cologne, Germany

skin lightening in East Asian populations (McEvoy et al. 2006; Norton et al. 2007).

The human *OCA2* gene (GenBank GeneID: 4948; accession numbers: NM_000275 and NT_000015), the homologue of the murine pink-eyed dilution (*p*) gene, plays a role in regulating the pH of melanosomes. Some mutations in the *OCA2* gene result in oculocutaneous albinism type II (OCA2). The *OCA2* gene spans 350 kb on chromosome 15q11.2–q12 and consists of 24 exons, encoding an 838-amino acid polypeptide (Lee et al. 1995; Oetting et al. 2005; Puri et al. 2000). A c.G1559A (Ala481Thr; unregistered in Entrez SNP) mutation in exon 14 was first discovered as a compound heterozygote in a European–American patient with apparent autosomal recessive ocular albinism, and its frequency was estimated to be 0.01 in the testing of 50 unrelated normal Caucasian subjects (Lee et al. 1994). Thereafter, this mutant allele, *OCA2*481Thr*, has sporadically been observed in Japanese OCA2 patients (Saitoh et al. 2000; Kato et al. 2003; Suzuki et al. 2003a; Kawai et al. 2005; Ito et al. 2006) and a German patient with congenital cataract and macular hypoplasia (Graw et al. 2006). A transfection study showed that *OCA2*481Thr* had approximately 70% function of the wild type allele in melanogenesis and confirmed it was a relatively mild OCA2 allele (Sviderskaya et al. 1997). This allele is not rare, but has been observed at a frequency as high as 0.10 in Japanese OCA patients and 0.12 in normally pigmented Japanese volunteers (Suzuki et al. 2003b), and it is warned that a number of subclinical patients of OCA2 with this allele might exist not only in Japan, but also all over the world (Kawai et al. 2005). However, as there have been only two studies on distribution of the A481T mutation, it remains unknown whether *OCA2*481Thr* exists in other populations. In this study, therefore, we investigated the frequency of *OCA2*481Thr* in more than 2,600 people from 20 African and Eurasian populations.

Subjects and methods

DNA samples were obtained from 2,615 unrelated individuals living in various areas of Eurasia (Table 1; Fig. 1). The donors were selected at random, irrespective of skin color, and their phenotypic data were unavailable. This study was approved by the Ethical Committee at the Faculty of Medicine, Tottori University. The typing of the A481T mutation was carried out based on the principle of the amplified product length polymorphism (APLP) method (Watanabe et al. 1997). The sequences of the primers used are as follows: OCA2-481-Thr: 5'-atatCACAAACATTGGAGGtGCTA-3' (nucleotide positions 115895–115914; lower cases are non-complementary nucle-

otides), OCA2-481-Ala: 5'-CACAAACATTGGAGGAaCTG-3' (115895–115914) and OCA2-481-R: 5'-TTGGAAACAATAATGACATTTGG-3' (115957–115935). The PCR cocktail consisted of 100 μ l of Multiple PCR Master Mix from a Multiplex PCR Kit (Qiagen), 20 μ l of three primers (1:3:1) with a concentration of 10 pmol/ μ l, and 80 μ l of water. PCR was performed in a volume of 8 μ l containing 7.5 μ l of the PCR cocktail and 0.5 μ l of a solution containing about 10 ng of genomic DNA. Cycle conditions were 95°C for 15 min, then 30 cycles of 94°C for 30 s, 56°C for 30 s, 72°C for 30 s, and a final extension step of 10 min at 72°C. The products were separated using a polyacrylamide gel (9%T, 5%C), then visualized by ethidium bromide staining. The homozygotes of *OCA2*481Thr* and *OCA2*481Ala* showed 67 and 63-bp bands, respectively, whereas the heterozygotes had both bands. Allele frequencies were estimated and Hardy–Weinberg equilibrium was tested using Arlequin program version 3.01 (Excoffier et al. 2005).

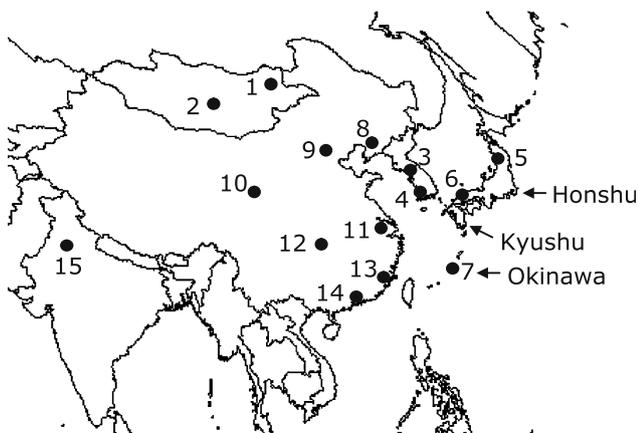
Results and discussion

Table 1 summarizes differences in the distribution of the A481T genotypes and allele frequency for the mutant in 20 populations. In each population, there was no statistically significant departure from Hardy–Weinberg equilibrium (data not shown). The highest value of the *OCA2*481Thr* frequency was observed in Buryat (0.24) and the second highest in Khalha (0.13). The frequencies in northern Han Chinese in Shenyang, Koreans, and Japanese were similar to each other and ranged from 0.075 in Tottori to 0.057 in Okinawa. These values are lower than the data (0.12) in Nagoya, central Japan (Suzuki et al. 2003). A slight but significant difference in distribution was observed between Okinawa and Nagoya ($\chi^2 = 3.92$, $P < 0.05$). In contrast, other Han Chinese populations were either low or zero. This allele was not found in Indo-European and African populations except for Turks in Germany. A recent study also failed to detect *OCA2*481Thr* in about 3,000 European descendants in Australia (Duffy et al. 2007). The frequency (0.01) in Caucasians reported previously (Lee et al. 1994) must have been overestimated. Thus, *OCA2*481Thr* is characteristic of northeastern Asian populations.

Buryat in Mongolia showed the highest frequency, but there may be some populations with higher frequencies in northeast China, Siberia, and/or Central Asia. Anyway, *OCA2*481Thr* is nearly restricted to a northeastern part of Asia. The distribution of *OCA2*481Thr* reflects the migration of ancient northeastern Asians. This allele must have occurred recently in a region of low ultraviolet radiation and have spread to northeast and Far East Asia. It

Table 1 Distribution of A481T Genotypes and *OCA2*481Thr* frequencies

No.	Population	Collection place	<i>n</i>	Genotypes of A481T			Frequency of <i>OCA2*481Thr</i>
				A	A/T	T	
1	Buryat	Dashbalbar	143	79	59	5	0.241
2	Khalha	Ulaan Baator	173	131	39	3	0.130
3	Korean	Seoul	139	120	19	0	0.068
4	Korean	Kwangju	141	121	19	1	0.074
5	Japanese	Yamagata	289	251	37	1	0.067
6	Japanese	Tottori	179	152	27	0	0.075
7	Japanese	Okinawa	87	78	8	1	0.057
8	Han Chinese	Shenyang	103	89	14	0	0.068
9	Han Chinese	Beijin	40	37	3	0	0.038
10	Han Chinese	Xi'an	109	101	8	0	0.037
11	Han Chinese	Wuxi	119	117	2	0	0.008
12	Han Chinese	Changsha	101	96	5	0	0.025
13	Han Chinese	Putien	118	112	6	0	0.025
14	Han Chinese	Huizhou	111	111	0	0	0.000
15	Indian	New Delhi	107	107	0	0	0.000
16	Turk	West Germany	200	195	5	0	0.013
17	German	West Germany	291	291	0	0	0.000
18	French	Rheims	98	98	0	0	0.000
19	Nigerian	West Germany	31	31	0	0	0.000
20	Ghanaian	West Germany	36	36	0	0	0.000

**Fig. 1** Location of Asian populations analyzed in the present study

showed a north–south downward geographical gradient. There is a significant correlation between allele frequencies and degrees of latitude in 14 East Asian populations ($r = 0.776$, $P < 0.01$). However, the frequencies are slightly higher in Korean and Japanese than in Han Chinese. This difference in distribution reminds us of the Y-chromosomal haplogroup C, frequencies of which differ remarkably between Han Chinese and northeast Asians including Buryat and Mongolian populations. Koreans and Japanese have higher frequencies than Han Chinese.

*OCA2*481Thr* has also made less contribution to the gene pools of Chinese Han populations than those of Far East populations, as haplogroup C could not expand extensively into mainland China due to the rapid and striking northward migration and expansion of haplogroup O from south China (Hammer et al. 2006; Xue et al. 2006). *OCA2*481Thr* has extended from northeast Asia through the Korean peninsula to the Japanese archipelago. The somewhat low frequency in Okinawa suggests a relatively weak gene flow from Kyushu to Okinawa. An east–west decreasing cline may also be observed in Eurasia. It is necessary to investigate the distribution in more detail and whether *OCA2*481Thr* in Europeans has the same origin as that in Asians. These expansions may also be a result of a certain selective advantage of hypofunction in pigmentation.

Individuals homozygous for *OCA2*481Thr* appear phenotypically normal (Suzuki et al. 2003b; Ito et al. 2006). However, this allele may affect variation in skin color in Japanese because of its hypofunction in pigmentation. Japanese skin type is classified on the basis of an individual's susceptibility to sunburn and ability to tan: type I, always burns, never tans; type II, moderately burns, moderately tans; type III, never burns, always tans (Sato and Kawada 1986). Subjects with type I showed statistically higher prevalence rates for actinic keratosis and nonmelanoma skin cancer in comparison with subjects

having the other types. The incidence of type I was estimated to be 18.6% in Kasai City, western Japan and 10.3% in Ie Island, Okinawa (Naruse et al. 1997; Nagano et al. 1999). These values coincide with total values of the homozygote and heterozygote for *OCA2*481Thr* in Honshu and Okinawa. Individuals with *OCA2*481Thr* may have less resistance to the stress of sunburn (Kawai et al. 2005).

Recently, mutant alleles in the melanocortin 1 receptor (*MC1R*), the membrane-associated transporter protein (*SLC45A2*), and a putative cation exchanger protein (*SLC24A5*) have been elucidated to play a key role in light skin and its variation in European populations (review, Sturm 2006). In contrast, little is understood about the genetic and molecular basis of light skin color in Asian populations. *OCA2*481Thr* described here is, of course, not a key mutation responsible for light skin pigmentation in Asians.

References

- Duffy DL, Montgomery GW, Chen W, Zhao ZZ, Le L, James MR, Hayward NK, Martin NG, Sturm RA (2007) A three-single-nucleotide polymorphism haplotype in intron 1 of *OCA2* explains most human eye-color variation. *Am J Hum Genet* 80:241–252
- Excoffier L, Laval G, Schneider S (2005) Arlequin ver 3.0: an integrated software package for population genetics data analysis. *Evol Bioinf Online* 1:47–50
- Graw J, Klopp N, Illig T, Preising MN, Lorenz B (2006) Congenital cataract and macular hypoplasia in humans associated with a de novo mutation in *CRYAA* and compound heterozygous mutations in *P*. *Graefes Arch Clin Exp Ophthalmol* 244:912–919
- Hammer MF, Karafet TM, Park H, Omoto K, Harihara S, Stoneking M, Horai S (2006) Dual origins of the Japanese: common ground for hunter-gatherer and farmer Y chromosomes. *J Hum Genet* 51:47–58
- Ito S, Suzuki T, Inagaki K, Suzuki N, Kono M, Tomita Y, Iwamoto T, Mochizuki N (2006) Two novel mutations detected in Japanese patients with oculocutaneous albinism. *J Dermatol Sci* 44:116–118
- Izagirre N, García I, Junquera C, de la Rúa C, Alonso S (2006) A scan for signature of positive selection in candidate loci for skin pigmentation in humans. *Mol Biol Evol* 23:1697–1706
- Jablonski NG, Chaplin G (2000) The evolution of skin coloration. *J Hum Evol* 39:57–106
- Kato A, Fukai K, Oiso N, Hosomi N, Saitoh S, Wada T, Shimizu H, Ishii M (2003) A novel *P* gene missense mutation in a Japanese patient with oculocutaneous albinism type II (*OCA2*). *J Dermatol Sci* 31:189–192
- Kawai M, Suzuki T, Ito S, Inagaki K, Suzuki N, Tomita Y (2005) A patient with subclinical oculocutaneous albinism type 2 diagnosed on getting severely sunburned. *Dermatology* 210:322–323
- Lao O, de Gruijter JM, van Duijn K, Navarro A, Kayser M (2007) Signatures of positive selection in genes associated with human skin pigmentation as revealed from analyses of single nucleotide polymorphisms. *Ann Hum Genet* 71:354–369
- Lee S-T, Nicholls RD, Bunday S, Laxova R, Musarella M, Spritz RA (1994) Mutations of the *P* gene in oculocutaneous albinism, ocular albinism, and Prader–Willi syndrome plus albinism. *N Engl J Med* 330:529–534
- Lee S-T, Nicholls RD, Jong MTC, Fukai K, Spritz AS (1995) Organization and sequence of the human *P* gene and identification of a new family of transport proteins. *Genomics* 26:354–363
- McEvoy B, Beleza S, Shriver MD (2006) The genetic architecture of normal variation in human pigmentation: an evolutionary perspective and model. *Hum Mol Genet* 15:R176–R181
- Myles S, Somel M, Tang K, Kelso J, Stoneking M (2007) Identifying genes underlying skin pigmentation differences among human populations. *Hum Genet* 120:613–621
- Nagano T, Ueda M, Suzuki T, Naruse K, Nakamura T, Taguchi M, Araki K, Nakagawa K, Nagai H, Hayashi K, Watanabe S, Ichihashi M (1999) Skin cancer screening in Okinawa, Japan. *J Dermatol Sci* 19:161–165
- Naruse K, Ueda M, Nagano T, Suzuki T, Harada S, Imaizumi K, Watanabe S, Ichihashi M (1997) Prevalence of actinic keratosis in Japan. *J Dermatol Sci* 15:183–187
- Norton HL, Kittles RA, Parra E, McKeigue P, Mao X, Cheng K, Canfield VA, Bradley DG, McEvoy B, Shriver MD (2007) Genetic evidence for the convergent evolution of light skin in Europeans and East Asians. *Mol Biol Evol* 24:710–722
- Oetting WS, Garrett SS, Brott M, King RA (2005) *P* gene mutations associated with oculocutaneous albinism type II (*OCA2*). *Hum Mutat* 25:323
- Puri N, Gardner JM, Brilliant MH (2000) Aberrant pH of melanosomes in pink-eyed dilution (*p*) mutant melanocytes. *J Invest Dermatol* 115:607–613
- Saitoh S, Oiso N, Wada T, Narazaki O, Fukai K (2000) Oculocutaneous albinism type 2 with a *P* gene missense mutation in a patient with Angelman syndrome. *J Med Genet* 37:392–394
- Satoh Y, Kawada A (1986) Action spectrum for melanin pigmentation to ultraviolet light, and Japanese skin typing. In: Fitzpatrick TB et al. (eds) *Brown melanoderma. Biology and disease of epidermal pigmentation*. University of Tokyo Press, Tokyo, pp 87–95
- Sturm RA (2006) A golden age of human pigmentation genetics. *Trends Genet* 22:464–468
- Suzuki T, Miyamura Y, Matsunaga J, Shimizu H, Kawachi Y, Ohyama N, Ishikawa O, Ishikawa T, Terao H, Tomita Y (2003a) Six novel *P* gene mutations and oculocutaneous albinism type 2 frequency in Japanese albino patients. *J Invest Dermatol* 120:781–783
- Suzuki T, Miyamura Y, Tomita Y (2003b) High frequency of the Ala481Thr mutation of the *P* gene in the Japanese population. *Am J Med Genet* 118A:402–403
- Sviderskaya EV, Bennett DC, Ho L, Bailin T, Lee ST, Spritz RA (1997) Complementation of hypopigmentation in *p*-mutant (pink-eyed dilution) mouse melanocytes by normal human *P* cDNA, and defective complementation by *OCA2* mutant sequences. *J Invest Dermatol* 108:30–34
- Watanabe G, Umetsu K, Yuasa I, Suzuki T (1997) Amplified product length polymorphism (APLP): a novel strategy for genotyping the ABO blood group. *Hum Genet* 99:34–37
- Xue Y, Zerjal T, Bao W, Zhu S, Shu Q, Xu J, Du R, Fu S, Li P, Hurles ME, Yang H, Tyler-Smith C (2006) Male demography in East Asia: a north–south contrast in human population expansion times. *Genetics* 172:2431–2439